

Kudzu root and maqui berry extract preclinical assessment in streptozotocin-induced nephropathy

Anup Patil¹, Saroj Yadav², M. Janani³, N.S Disha⁴, Mahesh Kumar Posa⁵, Saurabh Mishra⁶, I. Shaktheiswar⁷, Amit Kumar Singh^{8*}

¹Associate Professor, Department of Pharmacology Krishna Vishwa Vidyapeeth, Krishna Institute of Pharmacy, P. B Road, Malkapur, Karad. 415539, India

²Associate Professor, School of Medical and Allied Sciences, K.R. Mangalam University, Sohna Gurugram, Haryana, India

³Assistant Professor, Vel Tech High Tech Dr Rangarajan Dr. Sakunthala Engineering College Thiruvallur, Tamil Nadu, 600062, India

⁴Associate Professor, Department of Pharmaceutical Chemistry, RL Jalappa College of Pharmacy, Sree devaraj Urs Academy of Higher Education and Research (A Deemed to be University), Karnataka, India

⁵Associate Professor, School of Pharmaceutical Sciences/ Jaipur National University, Jaipur, Rajasthan, 302017, India

⁶Assistant Professor, College, Sitapur Sikhsha Sansthan, Sitapur, U.P., 261001, India

⁷Assistant Professor, Arulmigu Kalasalingam College of Pharmacy, Viruthunagar, Tamil Nadu, 626126, India

⁸Professor, Department of Pharmaceutics, United Institute of Pharmacy, (Affiliated to Dr. APJ Abdul Kalam Technical University, Lucknow), A-31/1, UPSIDC Industrial Area, Naini, Prayagraj -211010, Uttar Pradesh, India

***Corresponding author:** Amit Kumar Singh, Professor, Department of Pharmaceutics, United Institute of Pharmacy, (Affiliated to Dr. APJ Abdul Kalam Technical University, Lucknow), A-31/1, UPSIDC Industrial Area, Naini, Prayagraj -211010, Uttar Pradesh, India

Cite this paper as: Anup Patil, Saroj Yadav, M. Janani, N.S Disha, Mahesh Kumar Posa, Saurabh Mishra, I. Shaktheiswar, Amit Kumar Singh (2024) Kudzu root and maqui berry extract preclinical assessment in streptozotocin-induced nephropathy *.Frontiersin Health Informatics, 13 (6) 45-52*

ABSTRACT:

Background and Objectives: A metabolic problem related to diabetes mellitus is hyperglycemia, which happens when the body doesn't make enough insulin. Nephropathy is a major capillary problem that can happen with diabetes. The purpose of this study is to find out how well Maqui berry and Kudzu root products can stop experimental mice from getting nephropathy caused by diabetes.

Methods: Streptozotocin (55 mg/kg) was given intraperitoneally to male Wistar rats that weighed 200–250 g to make them develop diabetic nephropathy. To check for nephritis biochemically, blood glucose, total protein, albumin, urea, uric acid, creatinine, and total bilirubin were measured in serum and urine. Along with membrane-bound ATPases, lipid peroxidation, reduced glutathione, and superoxide dismutase were used to measure signs of oxidative stress.

Results: Nephropathy rats were given extracts from kudzu roots and maqui berries by mouth for two weeks. The doses were 50 mg/kg orally for kudzu and 100 mg/kg orally for maqui. Rats with diabetic nephropathy had much higher levels of creatinine, albumin, total protein, total bile, uric acid, urea, and signs of oxidative stress like lipid peroxidation. On the other hand, their levels of superoxide dismutase and glutathione were much lower. Over the course of two weeks, treatment with 50 mg/kg and 100 mg/kg of Kudzu root extract and 50 mg/kg and 100 mg/kg of Maqui berry extract, either alone or together, changed the levels of oxidative stress markers, ATPases, and biomarkers of renal function, getting closer to normal levels.

Conclusion: Synergistic benefits in decreasing oxidative stress indicators and hyperglycemia, thereby alleviating renal injury, were shown when Kudzu root extract and Maqui berry extract were used together, according to current research.

Keywords: Streptozotocin, Kudzu root extract, Maqui berry, diabetic nephropathy.

INTRODUCTION:

The prevalence of diabetes among adults increased from 4.7% in 1980 to 8.5% in 2014, as reported in the World Health Report that was released on November 15, 2017. Nearly 1.6 million people lost their lives in 2015 as a direct result of the disease. The World Health Organization predicts that diabetes will rank as the sixth leading killer in 2011 [1-3]. Diabetes mellitus (DM) is projected to affect 693 million individuals globally by the year 2045. The most prevalent form of diabetes worldwide is type 2 diabetes mellitus (T2DM), which occurs when the body fails to effectively use insulin. According to reports, the most common major ailment in 2017 was chronic kidney failure [2-4].

The gradual deterioration of kidney function due to persistently elevated blood sugar levels is known as diabetic nephropathy (DN). Diabetic nephropathy is characterized by impaired kidney function [3-5]. With or without chronic renal insufficiency leading to end-stage renal disease (ESRD), these patients may experience early microalbuminuria, hyperfiltration, hyperperfusion, increased capillary permeability to macromolecules, and proteinuria [4-6].

One of the most prevalent complications of diabetes is diabetic nephropathy, which affects the blood vessels. Additionally, it is the primary driver of end-stage renal illness, which manifests itself in an imbalance of inflammation, mesangial development, renal basement membrane breakdown, oxidative damage, and so on [5-7].

The gradual decline in kidney function, elevated protein levels, interstitial fibrosis, and glomerular failure are the hallmarks of diabetic nephropathy. Diabetic nephropathy can result from a number of factors, including changes in metabolic processes, kidney hemodynamics, increased renin-angiotensin production, and inflammation. Issues with metabolism and blood circulation are likely the root causes of diabetic nephropathy (DN) [6-8]. Due to their numerous beneficial effects on health, flavonoids are an integral component of numerous nutraceutical, medicinal, cosmetic, and pharmaceutical products. Reason being, they eliminate free radicals, shield cells from harm, and decrease inflammation quite effectively [7-9].

The goal of this research is to determine whether or not extracts from the Kudzu root and the Maqui berry can prevent diabetic nephropathy in experimental mice.

MATERIALS AND METHODS:

Materials:

Drugs and chemicals:

Both the kudzu root extract and the maqui berry extract originated from the Indian state of Maharashtra. Streptozotocin (STZ) was obtained from Sigma Aldrich in Mumbai, India. All ingredients and diagnostic tests required conformed to established specifications.

Experimental animals:

Eight sets of six adult male Wistar rats, ranging in weight from 200 to 250 grams apiece, were utilized for the experiment. Separated from one another, the rats resided in polypropylene containers lined with rice husks. These animals were housed in standard laboratory settings in accordance with the guidelines established by the Committee for the Purpose of Control and Supervision of Experiments on Animals (CPCSEA). Water and regular lab food were readily available to the animals prior to the dietary modification [10-12].

Induction of DN using STZ:

To cause diabetic nephropathy, a single dose of streptozotocin (55 mg/kg) diluted in 0.2 ml of citrate buffer was given. An automated glucometer checked the blood glucose levels (BGL) 72 hours after STZ treatment, which proved the person had diabetes. The rats were watched for four weeks to see how the nephritis got worse [13-16].

Experimental design:

The rats were put into eight groups, with six rats in each group. Here are the names of the groups: Group I: a treatment that only used a vehicle on the control rat Group II: STZ was given to rats. 11 Rats in Group III were given kudzu root extract that had been diluted in pure water. Rats in Group IV were given kudzu root extract that had not been diluted. 11 In Group V, rats were given distilled water that had Maqui berry extract added to it. In Group VI, rats also got distilled water that had Maqui berry extract added to it [17-20].

Serum and urine biochemical estimation:

After the second week of treatment, each rat was kept in its own metabolic box for one day, during which time a urine sample was taken. Microcapillaries were used to take blood from the retroorbital nerve while the person was anesthetized with ether. High-speed centrifugation was then used to separate the serum. For different biochemical analyses, serum and pee samples were kept at -20°C. A glucometer was used to measure the blood glucose levels. To check for total protein, albumin, urea, uric acid, creatinine, and total bilirubin in blood and urine samples, commercial diagnostic tools were used [21-25].

Tissue homogenization:

At the end of the treatment, the rats were killed by cutting off their heads. The kidneys were washed in cold physiological saline after they were cut out. The amount of antioxidant enzymes in the kidney was measured. The kidney was carefully cut into small pieces that were then put in a 0.25M sugar solution that was kept cold. To make the kidney cells more uniform, a 10% w/v tris-hydrochloride buffer was used. One that cools down was used to spin the mixture at 0°C and 10,000 rpm for 15 minutes. The amounts of lipid peroxidation (LPO), superoxide dismutase (SOD), and reduced glutathione (GSH) were measured in the supernatant. To test Na⁺/K⁺ ATPase, a sedimentation-based method was used [26-29].

Assessing oxidative stress signals:

Assessment of lipid peroxidation (LPO):

After 2.0 ml of freshly made 10% w/TCA was added to the kidney homogenate liquid, the mixture was put in an ice bath for 15 minutes. Centrifugation was used to separate the residue. Two milliliters of freshly made TBA were mixed with two milliliters of the clear leftover solution. The solution that was made was heated in a bath of hot water for ten minutes. After that, it was quickly cooled for five minutes in ice. A 532 nm reading was used to compare the pink color that formed when TBA and MDA reacted with the blank. Malondialdehyde that is standard was used in different amounts. The numbers were shown as MDA in nanomoles per mg of protein [30-35].

Assessment of reduced glutathione (GSH):

The kidney purée was centrifuged after being deproteinized with a 20% TCA solution. The next step was to combine 0.25 mL of the supernatant with 2 mL of the DTNB solution. Using phosphate buffer, the remaining portion was reduced to 3 ml. It was at 412 nm that the subsequent golden hue became visible. Standard glutathione was prepared in varying concentrations. Magnesium per gram of protein was used to measure the quantity of GSH [34-40].

RESULTS:

Effect of KRE, MBE, and combination on BGL:

Blood glucose levels were significantly greater in diabetic nephropathy rats compared to normal rats that had not been administered STZ after 72 hours. This role remained unchanged throughout week four. Table 1 shows that blood glucose levels in DN rats decreased more after KRE treatment than after no treatment at all, although the decreases were less pronounced with MBE alone treatment [41-44].

Table 1: Combined BGL effects of Kudzu root and Maqui berry extract

Sr. No.	Groups	Blood glucose level (mg/dl)		
		72hr after STZ injection	Final at week 4	Final after 6th week
1.	I	84.12±3.147	85.55±3.784	80.11±3.142
2.	II	255.6±8.36	332.4±8.641	254.7±3.34
3.	III	256.2±5.48	340.8±6.458	255.1±5.42
4.	IV	256.3±3.78	481.9±3.147	255.2±3.45
5.	V	256.8±3.47	443.7±3.254	258.3±3.25
6.	VI	242.7±6.89	440.3±8.456	243.6±5.56
7.	VII	266.8±8.46	302.4±6.148	267.1±7.25

8.	VIII	445.4±18.22	443.8±5.143	426.3±11.33
----	------	-------------	-------------	-------------

The effects of kudzu root and maqui berry on hypertrophy index, kidney weight, and urine volume:

By the end of the sixth week, we had measured the hypertrophy index, renal weight, and urine volume. After 24 hours, the urine volume (ml) of DN rats was noticeably higher than that of control rats. But because their kidneys weren't functioning as well, there was a marked shift in their hypertrophy index and kidney weight. In comparison to the DN group, the rats administered Kudzu root extract, Maqui berry extract, or a combination of the two showed a marked decrease in pee volume. Both their hypertrophy index and excessive kidney weight went back to normal (Table 2).

Table 2: Volume, weight, and hypertrophy index of the kidneys as affected by extracts from Kudzu root and Maqui berries

Sr. No.	Groups	Urine Volume			Kidney weight
		0 Day	4 th week	6 th week	
1.	I	15.2±0.81	16.3±0.82	15.3±0.89	0.11±0.02
2.	II	19.2±0.64	20.3±0.65	20.1±0.25	1.81±0.04
3.	III	20.3±0.82	21.5±0.89	21.2±0.46	1.12±0.02
4.	IV	17.3±0.38	18.5±0.42	18.2±0.43	1.23±0.05
5.	V	19.2±0.89	20.4±0.90	20.1±0.90	1.34±0.05
6.	VI	18.5±0.82	19.6±0.91	19.6±0.78	1.66±0.06
7.	VII	19.2±0.14	20.3±0.23	20.4±0.26	1.18±0.05
8.	VIII	20.4±0.65	21.6±0.70	19.5±0.55	1.37±0.01

Hormonal and biochemical effects of Kudzu root and Maqui berry extracts on urine:

At the six-week mark, we re-analyzed the urine for albumin, urea, and total protein levels. Significant alterations occurred in the concentrations of albumin, urea, and total protein within the bodies of diabetic nephropathy rats. When compared to non-DN rats, DN rats' albumin, urea, and total protein levels were significantly greater. Kidney biomarker levels decreased significantly in DN rats treated with Kudzu root extract, Maqui berry extract, or a combination of the two. Serum creatinine, total bilirubin, and uric acid levels were assessed six hours later. Diabetic nephropathy rats' urine contained significantly greater concentrations of creatinine, total bilirubin, and uric acid compared to normal rats' urine. Rats' uric acid, creatinine, and total bilirubin levels were dramatically altered after administration of Kudzu root extract, Maqui berry extract, or a combination of the two. Table 3 shows that after the second week of therapy, all treatment groups had significantly lower biomarker levels than the DN group.

Table 3: Hormonal and biochemical effects of Kudzu root and Maqui berry extracts on urine

Sr. No.	Groups	Total Protein	Albumin	Urea	Creatinine	Bilirubin
1.	I	4.35±0.06	1.23±0.03	16.3±0.14	1.23±0.01	0.87±0.03
2.	II	8.29±0.08	4.29±0.02	21.1±0.26	3.29±0.03	6.45±0.05
3.	III	6.31±0.06	3.31±0.05	22.2±0.47	4.31±0.04	2.12±0.01
4.	IV	8.56±0.08	4.56±0.07	35.2±0.45	5.56±0.06	3.23±0.04
5.	V	6.21±0.09	4.21±0.08	35.1±0.84	3.21±0.07	2.34±0.06
6.	VI	5.58±0.06	3.58±0.04	35.6±0.80	2.58±0.05	2.66±0.04
7.	VII	5.46±0.04	4.46±0.05	23.4±0.30	4.46±0.04	2.18±0.06
8.	VIII	5.75±0.05	2.75±0.06	23.5±0.61	3.75±0.05	3.37±0.02

Metabolic and enzymatic effects of Kudzu root and Maqui berry extracts on blood samples:

Blood samples were taken after six weeks to measure albumin, urea, and total protein levels. Rats suffering from diabetic nephropathy showed substantial changes in albumin, urea, and total protein concentrations. The levels of creatinine, total bilirubin, and uric acid were noticeably greater ($p<0.001$) in the urine of rats with diabetic nephropathy compared to normal rats. After six weeks, there were notable variations in the blood levels of uric acid, creatinine, and total bilirubin among rats that were administered Kudzu root extract, Maqui berry extract, or a combination of the two. Rats with

diabetic nephropathy had significantly higher concentrations of creatinine, total bilirubin, and uric acid in their urine compared to rats without the condition. Rats' uric acid, creatinine, and total bilirubin levels were dramatically altered after administration of Kudzu root extract, Maqui berry extract, or a combination of the two [45-48]. Table 4 shows that after two weeks of treatment, all treatment groups had significantly lower biomarker levels than the DN group.

Table 4: The impact on serum biochemical parameters of extracts from Kudzu root and Maqui berries, as well as their combination

Sr. No.	Groups	Total Protein	Albumin	Urea	Creatinine	Bilirubin
1.	I	5.35±0.05	2.23±0.03	20.3±0.14	1.23±0.01	0.88±0.06
2.	II	13.29±0.08	5.29±0.02	52.1±0.26	3.29±0.03	4.45±0.04
3.	III	4.31±0.04	4.31±0.05	33.2±0.47	4.31±0.04	3.12±0.02
4.	IV	5.56±0.07	3.56±0.07	36.2±0.45	5.56±0.06	4.23±0.03
5.	V	4.21±0.08	6.21±0.08	38.1±0.84	3.21±0.07	1.34±0.04
6.	VI	3.58±0.05	2.58±0.04	31.6±0.80	2.58±0.05	3.66±0.03
7.	VII	4.46±0.04	3.46±0.05	29.4±0.30	4.46±0.04	4.18±0.04
8.	VIII	3.75±0.08	1.75±0.06	31.5±0.61	3.75±0.05	2.37±0.03

CONCLUSION:

Natural antioxidants like maqui berry and kudzu root extract may help manage high blood sugar and the oxidative damage that comes with diabetes, according to a recent study. The kidneys were strongly protected from damage when treated with kudzu root extract and maqui berry extract, which enhanced the efficiency of the antioxidant system and corrected morphological alterations. Compared to using just one of the antioxidants, the combined effect was much stronger when all three were employed. The study found that diabetic nephropathy was able to be prevented from worsening by combining the strong antioxidant properties of kudzu root extract with those of maqui berry extract.

Funding:

None

Conflict of Interest:

None

REFERENCES:

- Feinberg T, Wieland LS, Miller LE, Munir K, Pollin TI, Shuldiner AR, Amoils S, Gallagher L, Bahr-Robertson M, D'Adamo CR. Polyherbal dietary supplementation for prediabetic adults: study protocol for a randomized controlled trial. *Trials*. 2019 Dec;20:1-3.
- Dinda B, Dinda S, Chakraborty M. Pharmacology of Anti-Obesity and Antidiabetic Phytochemicals Isolated from Various Natural Sources (Plants, Seaweeds, Mushrooms, Marine Animals, and Microorganisms). In *Natural Products in Obesity and Diabetes: Therapeutic Potential and Role in Prevention and Treatment* 2022 Mar 9 (pp. 277-467). Cham: Springer International Publishing.
- Zhu J, Xing G, Shen T, Xu G, Peng Y, Rao J, Shi R. Postprandial Glucose Levels Are Better Associated with the Risk Factors for Diabetes Compared to Fasting Glucose and Glycosylated Hemoglobin (HbA1c) Levels in Elderly Prediabetics: Beneficial Effects of Polyherbal Supplements—A Randomized, Double-Blind, Placebo Controlled Trial. *Evidence-Based Complementary and Alternative Medicine*. 2019;2019(1):7923732.
- Liu D, Wen Q, Liu M, Gao Y, Luo L, Zhang Z, Chen Q. Dietary supplements for prediabetes: A protocol for a systematic review and meta-analysis. *Medicine*. 2020 May 15;99(20):e20347.
- Keservani RK, Sharma AK, Kesharwani RK, IGI G. *Phytopharmaceuticals and its applications in therapy. Recent Advances in Drug Delivery Technology*. 2017:202-29.
- Kiani Z, Amini S, Askari G, Kesharwani P, Bagherniya M, Sahebkar A. The effect of phytochemicals in prediabetic patients: A systematic review of randomized controlled trials. *Phytotherapy Research*. 2023 Aug;37(8):3239-61.
- Ahmad, Mohd F., Ahmad, S. M., Keservani, Raj K., Sharma, Anil K. A Study on Anti-Inflammatory Activity

- of Tuber Extracts of *Solanum Tuberosum* (Solanaceae) in Male Albino Rats, *Nat. Aca. Sci. Lett. India*. 2016, 39 (6): 421-425.
8. Keservani, Raj K., Sharma, Anil K., Jarouliya, U., Singh, A.K. Ebola Virus Disease and its Complications *Universal Journal of Pharmaceutical Research*, 2016c, 1 (1), 54-58.
 9. Keservani, Raj K., Kesharwani, Rajesh K., Sharma, Anil K., Naturally Occurring Toxicants as Etiologic Agents of Food-borne Disease, In: *Food Toxicology*, Edited by Bagchi, Debasis., Swaroop, Anand, CRC Press, Taylor and Francis. Chapter 12, 2016d, 245-262. ISBN: 9781498708746
 10. Ahmad, Mohd F., Ahmad, S. M., Keservani, Raj K., Pradhan, A. "Anti-Ulcer Activity of Tuber Extracts of *Solanum Tuberosum* (Solanaceae) In Rats", *Acta Fac. Pharm. Univ. Comen. LXII*, 2015a (2): 32-37.
 11. Jarouliya U, Zacharia A, Keservani RK, Prasad GB. Spirulina maxima and its effect on antioxidant activity in fructose induced oxidative stress with histopathological observations. *European Pharmaceutical Journal*. 2015;62(2):13-9.
 12. Ahmad, Mohd F., Keservani, Raj K., Babu, DJM., Interaction study between *Ocimum sanctum* and Glimepiride (sulfonylurea derivative) in diabetic rats, *J. Chin. Pharm. Sci.* 2015b, 24 (3), 156–163.
 13. Keservani Raj K., Shelke Suvarna Jagannath, Gawali Vikas, Gaviraj E.N., Binorkar Sandeep V., Rane Sachin Shrikrushna, Sarvadnya Anagha Amit and Patil Sharangouda J.: Anti-Alzheimer effect of *Ammannia baccifera* whole plants ethanolic extract. *Intern. J. Zool. Invest.* 2024, 10(2): 671-678.
 14. Arya RK, Sati D, Bisht D, Keservani RK. Nanotechnology-Based Bacterial Immunotherapy. In *Nutraceuticals and Functional Foods in Immunomodulators 2023* Jan 1 (pp. 3-19). Singapore: Springer Nature Singapore.
 15. Keservani RK, Sharma AK. Immunomodulatory and Antimicrobial Effects of Nanoemulsions: Pharmaceutical Development Aspects and Perspectives on Clinical Treatments. In *Nanodispersions for Drug Delivery 2018* Sep 24 (pp. 115-130). Apple Academic Press.
 16. Lim WX, Gammon CS, von Hurst P, Chepulis L, Page RA. A narrative review of human clinical trials on the impact of phenolic-rich plant extracts on prediabetes and its subgroups. *Nutrients*. 2021 Oct 22;13(11):3733.
 17. Talele SG, Ahire ED, Talele GS, Derle DV. An innovative approach as self-emulsifying drug delivery system for phytoconstituents. In *Enhancing the Therapeutic Efficacy of Herbal Formulations 2021* (pp. 69-84). IGI Global.
 18. Gautam T, Gautam SP, Keservani RK, Sharma AK. Phytochemical screening and wound healing potential of *Cuscuta reflexa*. *Journal of Chinese Pharmaceutical Sciences*. 2015 May;5:003.
 19. Yadav S, Upadhyay KK, Praveena R., Dharmalingam SR, Sulthana N, Singh AK, Dharmalingam S, & Keservani RK,. (2024). Formulation, Evaluation, And In Vitro Antifungal Assessment of A Topical Preparation Using Leaf Extract From *Tridax Procumbens* L. *Revista Electronica De Veterinaria*, 25(1), 2323 -2327. <https://doi.org/10.69980/redvet.v25i1.1185>
 20. Pulavarthy Vishnu, Manoj Gangadhar Shinde, Anand Mundada, Samiksha Dilipkumar Jayswal, Tilotma Sahu, Sudhahar Dharmalingam, Sangeetha Menon, & Raj K. Keservani. (2024). Molecular Docking Study Of Some Phytoconstituents Of *Alangium Salviifolium* Against Colorectal Cancer. *Revista Electronica De Veterinaria*, 25(1S), 1308 -1314. <https://doi.org/10.69980/redvet.v25i1S.1140>
 21. Ranjith K., Pulusu V.S., Keservani Raj K., Kachave R.N., Anumolu D.P., Tiwari N., Lohani, H., & Mundada A., (2024). Method Development & Validation of Stability Indicating Assay Method of Clindamycin in Adapalene and Clindamycin Gel Formulation by RP-HPLC. *Revista Electronica De Veterinaria*, 25(1S), 1247 - 1259. <https://doi.org/10.69980/redvet.v25i1S.1100>
 22. Uvaraja V.C., Keservani R.K., Maurya NK., Pendakur B., and Adhoni SA., (2024) Formulation and development of gel with essential oils and effect of polymer on their antimicrobial activity. *Biochem. Cell. Arch.* 24, 0000-0000. DOI: <https://doi.org/10.51470/bca.2024.24.2.0000>
 23. Uvaraja V.C., Binorkar Sandeep V., Parlikar Gajanan R., Pallavi Runjhun., S Renuka Jyothi., Keservani Raj K., N Kale Ramdas., Haleshappa R. Evaluation Of MT-Liver Protective and Antioxidant Effects Of Red Algae, *Nanotechnology Perceptions* 20 No. S11 (2024) 1264-1272.
 24. Suvarna R, Shenoy RP, Hadapad BS, Nayak AV. Effectiveness of polyherbal formulations for the treatment of type 2 Diabetes mellitus-A systematic review and meta-analysis. *Journal of Ayurveda and integrative medicine*. 2021 Jan 1;12(1):213-22.
 25. Vitetta L, Butcher B, Dal Forno S, Vitetta G, Nikov T, Hall S, Steels E. A Double-Blind Randomized Placebo-

- Controlled Study Assessing the Safety, Tolerability and Efficacy of a Herbal Medicine Containing Pycnogenol Combined with Papain and Aloe vera in the Prevention and Management of Pre-Diabetes. *Medicines*. 2020 Apr 22;7(4):22.
26. Sable RR, Thite DG, Udavant PB, Ahire ED, Khairnar SJ. Isolation and identification of flavonoids compounds and formulation of tecoma undulata (Sm.) seem. Herbal ointment. *Materials Today: Proceedings*. 2023 Dec 12.
27. Venkatakrishnan K, Chiu HF, Wang CK. Popular functional foods and herbs for the management of type-2-diabetes mellitus: A comprehensive review with special reference to clinical trials and its proposed mechanism. *Journal of Functional Foods*. 2019 Jun 1;57:425-38.
28. Siddiqui SA, Khan S, Wani SA. Controlling diabetes with the aid of medicinal herbs: A critical compilation of a decade of research. *Critical Reviews in Food Science and Nutrition*. 2023 Dec 20;63(33):12552-66.
29. Nakanekar A, Kohli K, Tatke P. Ayurvedic polyherbal combination (PDBT) for prediabetes: A randomized double blind placebo controlled study. *Journal of Ayurveda and integrative medicine*. 2019 Oct 1;10(4):284-9.
30. Ghorbani A. Clinical and experimental studies on polyherbal formulations for diabetes: current status and future prospective. *Journal of integrative medicine*. 2014 Jul 1;12(4):336-45.
31. Ghosh S, Chattopadhyay B, Koley M, Maiti S, Gourav K, Gupta S, Saha S. Plant extracts as add-on therapeutics in homeopathy: An open-label, randomized trial using mother tinctures in prediabetes. *Journal of Integrative and Complementary Medicine*. 2022 Sep 1;28(9):757-67.
32. Shane-McWhorter L. Botanical dietary supplements and the treatment of diabetes: what is the evidence?. *Current Diabetes Reports*. 2005 Sep;5(5):391-8.
33. Derosa G, D'Angelo A, Maffioli P. The role of selected nutraceuticals in management of prediabetes and diabetes: An updated review of the literature. *Phytotherapy Research*. 2022 Oct;36(10):3709-65.
34. Saad B, Zaid H, Shanak S, Kadan S, Saad B, Zaid H, Shanak S, Kadan S. Hypoglycemic and Anti-obesity Polyherbal Mixtures. *Anti-diabetes and Anti-obesity Medicinal Plants and Phytochemicals: Safety, Efficacy, and Action Mechanisms*. 2017:217-51.
35. Chen J, Jin L, Chen M, Xu K, Huang Q, He B. Application of natural compounds in the treatment and prevention of prediabetes. *Frontiers in Nutrition*. 2023 Nov 30;10:1301129.
36. Adeoye OB, Iyanda AA, Daniyan MO, Adeoye DA, Olajide OL, Akinlawo OO, Adetunji AO, OsundinA BO, Olatinwo OM. Anti-dyslipidaemia and cardio-protective effects of Nigerian bitter honey in streptozotocin induced diabetic rats. *Universal Journal of Pharmaceutical Research* 2023; 8(2):10-18. <https://doi.org/10.22270/ujpr.v8i2.920>
37. Mirza MS, Ali SA, Sanghvi I. Evaluation of methanolic extract of Euphorbia neriifoliastem bark on blood sugar levels, serum and tissue lipids in a preclinical model. *Universal Journal of Pharmaceutical Research* 2017;2(3): 1-5. <http://doi.org/10.22270/ujpr.v2i3.R1>
38. Kowa TK, Nyemb N, Tchamgoue AD, Tabi TGN. Antihyperglycemic and anti-oxidant potential of ethanol extract of Vitex thyrifloraleaves on diabetic rats. *Universal Journal of Pharmaceutical Research* 2018; 3(3): 19-24. <https://doi.org/10.22270/ujpr.v3i3.16>
39. Hou K, Chen C, Wang X, Guo Y. Effects of emodin on blood glucose and body weight in type 1 diabetic rats. *Universal Journal of Pharmaceutical Research* 2017; 2(4): 11-14. <http://doi.org/10.22270/ujpr.v2i4.R3>
40. Al-Kaf AG, Nelson NO, Patrick OU, Peace AN, Victor EJ, Okolie SO, Alexander I. Phytochemical screening and antidiabetic activity of methanolic extract of Caylusea abyssinicalleaves. *Universal Journal of Pharmaceutical Research* 2022; 7(6):39-45. <https://doi.org/10.22270/ujpr.v7i6.868>
41. Al-Baoqai N, Al-Mahbashi H, Al-Adhal A. Antidiabetic and antihyperlipidemic activity of Dracaena cinnabariBalf. Resin ethanolic extract of Soqatra Island in Experimental Animals. *Universal Journal of Pharmaceutical Research* 2018; 3(5): 1-10. <https://doi.org/10.22270/ujpr.v3i5.19>
42. Mahadeva Rao US. Phytochemical screening and in vitro antioxidant and anti-diabetic potentials of Persea americanamill. (Lauraceae) fruit extract. *Universal Journal of Pharmaceutical Research* 2018; 3(5): 34-41. <https://doi.org/10.22270/ujpr.v3i5.200>
43. Idoko A, Emmanuel UEG, Catherine OI. Phytochemical screening of aqueous, ethanol and methanol extracts of Flacourtia indicaleaf and ripe fruit. *Universal Journal of Pharma-ceutical Research* 2022; 7(5):18-22. <https://doi.org/10.22270/ujpr.v7i5.83>
44. Dinesh S, Sharma S, Chourasiya R. Therapeutic Applications of Plant and Nutraceutical-Based Compounds for

- the Management of Type 2 Diabetes Mellitus: A Narrative Review. *Current Diabetes Reviews*. 2024 Feb 1;20(2):112-30.
45. Yeh GY, Eisenberg DM, Kaptchuk TJ, Phillips RS. Systematic review of herbs and dietary supplements for glycemic control in diabetes. *Diabetes care*. 2003 Apr 1;26(4):1277-94.
46. Zaid H, Saad B, Mahdi AA, Tamrakar AK, Haddad PS, Afifi FU. Medicinal plants and natural active compounds for diabetes and/or obesity treatment. *Evidence-based complementary and alternative medicine: eCAM*. 2015;2015.
47. Heydari M, Hashempur MH, Daneshfard B, Mosavat SH. Bioactive foods as dietary intervention for diabetes from the perspective of Persian medicine. In *Bioactive Food as Dietary Interventions for Diabetes* 2019 Jan 1 (pp. 49-68). Academic Press.
48. Cefalu WT, Ye J, Wang ZQ. Efficacy of dietary supplementation with botanicals on carbohydrate metabolism in humans. *Endocrine, Metabolic & Immune Disorders-Drug Targets (Formerly Current Drug Targets-Immune, Endocrine & Metabolic Disorders)*. 2008 Jun 1;8(2):78-81.