

Evaluation of Phytochemical Screening Anti-Convulsant Activity of Ethanolic Extract of *Plumbago Zeylanica*

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ABSTRACT

The purpose of this study is to explore the anticonvulsant effect of the leaf gall extract of *Plumbago zeylanica* using pentylene tetrazole-induced convulsion (PTZ) in rats. *Plumbago zeylanica* is an indigenous plant belonging to the family Fabaceae (Papilionaceae) commonly known as Karanj. Freshly powdered flowers were extracted with 70% ethanol. The convulsion is induced by the administration of pentylene tetrazole (80 mg/kg, i.p.) to wistar albino rats, and those showing response was divided into three groups of six animals each. The group I treated with 1% normal saline (1ml/100gm, orally), Groups II treated with phenytoin sodium (25 mg/kg, i.p.) and Groups III treated with ethanolic extract at a dose of (250 mg/kg, i.p.). The ethanolic extract showed significant anticonvulsant activity by lowering the duration of the extension phase (3.72 ± 0.65) when compared to the control group (8.94 ± 0.42). From the experiment, we can say *Plumbago zeylanica* had significant anticonvulsant activity.

KEYWORDS: *Plumbago zeylanica* Phenytoin sodium, Pentylene tetrazole induced convulsion, anticonvulsant activity

INTRODUCTION

Plumbago zeylanica is an [herbaceous plant](#) with glabrous stems that climb prostrate, or erect. The leaves are petiolate or sessile and have ovate, lance-elliptic, or spatulate to oblanceolate blades that measure $5-9 \times 2.5-4$ cm in length. Bases are attenuated while apexes are acute, acuminate, or obtuse. Inflorescences are 3–15 cm long, and have glandular, viscid rachises. Bracts are lanceolate and $3-7 \times 1-2$ mm long. The heterostylous flowers have white corollas 17–33 mm in diameter and tubes 12.5–28 mm in length. Capsules are 7.5–8 mm long and contain reddish-brown to dark-brown seeds.¹ *Plumbago zeylanica* L. (Synonym: *P. viscosa* Blanco)

(chromosome number $2n=24$) is a multipurpose medicinal herb of the family Plumbaginaceae. A native of South Asia, the species is distributed throughout most of the tropics and subtropics; growing in deciduous woodland, savannas' and scrublands from sea-level up to 2000 m altitude^{2,5}

It has been recognized in different systems of traditional medicines for the treatment of different diseases and ailments of humans⁶. It contains several phytoconstituents belonging to category flavonoids and fixed oils. These candidates show antimicrobial and anthelmintic activity^{7,8}.

Leaf galls are a disturbing sight but are not usually as serious as they appear. These bumps and deformities are generally the result of feeding insects or some other foreign organism such as bacteria, fungi, mites, nematodes, and even viruses. The bumps are generally not caused by a disease. They are usually insect damage. For galling aphids and their hosts, tannins are crucial for plant-insect interactions and for protecting the host plant from herbivory. Due to their peculiar chemical characteristics, tannins from plant galls have been used for medical and chemical purposes for more than 2000 years. In this study, hydrolyzable tannin concentrations in galls increased from gall initiation (38.34% on June 21) to maturation (74.79% on August 8), then decreased gradually thereafter (58.83% on October 12). A total of 81 genes (named as GTS1-81) with putative roles in Gallo tannin biosynthesis and 22 genes (TS1-22) in condensed tannin biosynthesis. Multiple genes encoding 1-Beta-D- glucosyl transferases may play a vital role in Gallo tannin accumulation in plant galls⁹. Crown galls form as a result of wound inoculation on many species of plants by the soil-dwelling bacterium *Agrobacterium tumefaciens*.

Tannins are water-soluble polyphenols that are present in many plants and these are more abundant in vulnerable parts of plants, such as new leaves and flowers¹⁰. In all vascular plants studied, tannins are manufactured by a chloroplast-derived organelle, the tannosome. Tannins are mainly physically located in the vacuoles or surface wax of plants. The two most important components of tannins are tannic acid and Gallic acid. Tannic acid is widely used in therapeutics, and it is derived from galls on the leaves, which are the result of parasitic insect activity. They can be effective in the treatment of various diseases by the multitude of their beneficial activities such as anti-inflammatory¹¹, anti-oxidative¹², anti-convulsant¹³, and antitumor behaviors¹⁴. The pivotal neuroprotective effects of tannins are related to their capacity to act as free radical scavengers and to activate the antioxidant system in the body, thereby including protection against neurotoxins and oxidative-stress-induced neuronal damage, neuronal inflammation, and some other factors.

The flowers and sprouts of *Plumbago zeylanica* were used in folk remedies for tumors¹⁵. *Plumbago zeylanica*, commonly known as Ceylon leadwort, doctor bush or wild leadwort is a species of *Plumbago* with a [pantropical](#) distribution. Seed extract of this plant has hypotensive effects and produces uterine contractions. Powdered seed is used in bronchitis, chronic fever, whooping cough, and chronic skin diseases and painful rheumatic joints¹⁶. Leaves are active against *Micrococcus*; their juice is used for cold, cough, diarrhea, dyspepsia, flatulence, gonorrhea, and leprosy¹⁷. Roots are used for cleaning gums, teeth and ulcer. *Plumbago zeylanica* plant is used for anti-inflammatory¹⁸, anti-plasmodial¹⁹, anti-nociceptive²⁰, anti-hyperglycemic, anti-lipid oxidative, antidiarrheal, anti-ulcer, anti-hyperammonic, CNS depressant activity²¹ and antioxidant activity.

2. MATERIALS AND METHODS

2.1 Collection of Plant material

Collection and authentication of plant material The leaf galls of *Plumbago zeylanica* was collected from Maruthi Nagar, Guntur, Andhra Pradesh, India and material was identified and authenticated by Dr. B.R.C Murthy, reader in Botany, Hindu college, Guntur, India.

Preparation of test extracts: Crushed powders of species were successively soxhlet extracted. Later, each of the homogenates was filtered and the residue was re-extracted twice for complete exhaustion, the extracts were cooled individually. Each filtrate was concentrated to dryness in vitro and re-dissolved in respective solvents, were stored at 4°C in a refrigerator, until screened for phytochemical activity. Phytochemical Screening: Phytochemical screening was performed using standard procedure: Test for Reducing sugar (Fehling's Test): The aqueous extract (0.5gm in 5 ml of water) was added to boiling Fehling's solution (A and B) in a test tube. The solution was observed for a color reaction²².

Test for Flavonoids: 4ml of extract solution was treated with 1.5ml of 50% methanol solution. The solution was

warmed and metal magnesium was added. To this solution, 5-6 drops of concentrated Hydrochloride acid were added and red color was observed for flavonoids and orange color for flavones²³.

Test for Alkaloids: Alkaloids solutions produce white yellowish precipitate when a few drops of Mayer's reagents are added. Most alkaloids are precipitated from neutral or slightly acidic solution by Mayer's reagent. The alcoholic extract was heated in a boiling water bath with 2% hydrochloric acid. After cooling, the mixture was filtered and treated with a few drops of Mayer's reagent. The sample was then observed for turbidity or yellow precipitation²³.

Test for Tannins: About 0.5 g of extract was boiled in 10 ml of water in a test tube and then filtered. A few drops of 0.1% ferric chloride were added and observed for brownish green or a blue-black coloration²⁴.

Test for Terpenoids (Salkowski Test): To 0.5 gm each of the extracts was added to 2ml of chloroform. Concentrated sulphuric acid (3ml) was carefully added to form a layer. Reddish-brown coloration of the interface indicates the presence of terpenoids²⁵.

Test for carbohydrates: Phytochemical screening *Plumbago zeylanicum* shows that the plant has abundant amount of Secondary metabolites in it. The following table shows the results of tests performed. Preliminary phytochemical screening of the plant *Plumbago zeylanica* leaves (chitrak) [(+: present) (-: absent)]²⁶.

Preparation of *Plumbago zeylanica* extract

After collection, the leaf galls and flowers were shade-dried, pulverized using an electrical grinder and passed through the sieve (40 mesh size). The extract was prepared cold maceration method by taking 100g of powder in ethanol and soaked about 7 days. The extract was filtered using muslin cloth, followed by Whatman filter paper (No.1) and concentrated at reduced temperature on rotary evaporator. The extract was evaporated to dryness to achieve the yield of 8% (w/w) of the extract and kept in a desiccator²⁷. We made experiments to collect the fraction of extract using column chromatography with different solvents of varying polarity and finally the ethanolic fraction was selected for the study as it has given number of colored bands (Table 1). The 39th fraction (C-39) of ethanolic extract was collected and used for the evaluation of anti-convulsant activity.

Table no :1 Phytochemical screening of leaf gall and flower extract of *Plumbago zeylanica*.

Plant Extract	Reducin g sugar	Flavonoid s	Alkaloid s	Glycoside s	Tannin s	Terpenoid s	Carbohydrate s	Saponin s
Distilled Water	+ve	+ve	+ve	+ve	-ve	+ve	+ve	+ve
Ethyl acetate	+ve	+ve	+ve	+ve	+ve	+ve	+ve	+ve
Pet.ether	+ve	+ve	+ve	-ve	+ve	+ve	+ve	-ve
Benzene	+ve	-ve	-ve	-ve	-ve	+ve	-ve	-ve
Methanol	+ve	-ve	-ve	+ve	+ve	-ve	+ve	+ve
Chloroform	+ve	+ve	-ve	-ve	+ve	-ve	+ve	-ve

2.3. Experimental Animals

Animals Five male white albino Wistar rats weighing 150-200g were selected and procured from the animal house of Hindu college of Pharmacy, Guntur. The animals were maintained in well-cages. They were fed with a rodent diet and water ad libitum. Animal experiments were performed according to the ethical guidelines suggested by the Institutional Animal Ethics Committee (IAEC) and guided by the Committee for Control and Supervision of Experiments on Animals (CPCSEA) with registration number 1263/PO/ac/09/CPCSEA, dated 05/05/2014. All efforts were made to minimize animal suffering and to reduce the number of animals used. The Institutional animal ethical committee has approved the protocol of the study²⁸.

2.4. Experimental design

The anticonvulsant activity of *Plumbago zeylanica* flower extract was evaluated for pentylene tetrazole-induced convulsion (PTZ) in rats¹³. The convulsion is induced by administration of pentylene tetrazole (80 mg/kg, i.p.) to Wistar albino rats. Rats showing responses were divided into three groups of six animals each. The first group of animals was administered 1% normal saline (1ml/100gm) orally which served as a negative control. Group of II animals were treated with phenytoin sodium (obtained from Merck, India) (25 mg/kg, i.p.) which served as positive control¹⁴. Group of III animals (Ps were treated with ethanolic extract of PPLE at a dose of (250 mg/kg i.p.). Drug pretreatment was given 1 hr the administration of Pentylenetetrazole and each animal was placed in an individual plastic cage and observed initially 30 min. and later up to 24 hrs for the duration of tonic, flexion, tonic extension, clonus, and death/recovery²⁹. Statistical analysis Statistical analysis was done using a one-way analysis of variance (ANOVA) followed by a Student's t-test. Results are expressed as means \pm SEM from six rats in each group³⁰⁻³².

3. RESULTS

Plumbago zeylanica flower extract was subjected for an anticonvulsant effect using PTZ induced convulsion model in rats. PPLE exhibits significant anticonvulsant activity by lowering the duration of the extension phase when compared to the control group. The duration of tonic and hind limb extension in rats with 70% ethanolic extract was 3.72 ± 0.65 at a dose of 250 mg/kg. The activity of ethanolic extract was comparable ($P > 0.001$) to that produced by standard drug phenytoin sodium (Table 2).

Table 2. Anticonvulsant activity *Plumbago zeylanica* flower extract on Pentylenetetrazole induced convulsion in rats.

Treatment	Dose	Time in seconds in various phases of flexion and extension				
		Flexion	Extension	Convulsion	Stupor	Recovery/ death
Control (normal saline)	100 mg /1 ml	3.83 ± 0.36	3.83 ± 0.368	4.65 ± 0.71	104.10 ± 8.74	Recovery
Phenytoin sodium	25 mg/Kg	2.15 ± 0.21	9.41 ± 0.42	7.86 ± 0.35	91.55 ± 5.85	Recovery
Ethanolic extract	250 mg	1.90 ± 0.25	3.72 ± 0.6	8.96 ± 2.00	102 ± 8.75	Recovery

Data are expressed in mean \pm SEM, n = 6 in each group *P < 0.001 vs Phenytoin sodium-treated rats by student's t-test

4. DISCUSSION

Pentylenetetrazole is a selective blocker of the chloride ionophore complex to the GABA-A receptor, and after repeated or single dose administration leads to a decrease in GABAergic function and to the stimulation and modification of density or sensitivity of different glutamate receptor subtypes in many brain regions. Pentylenetetrazole may also triggers various biochemical processes including the activation of the membrane phospholipase, proteases, and nucleases. Alteration in membrane phospholipids metabolism causes liberation of free fatty acids, diacylglycerols, eicosanoids, lipid peroxidase, and free radicals. The tonic extensor phase is selectively abolished by the drugs effective in generalized tonic-clonic seizure¹¹. Phytochemicals such as kaempferol and fixed oils are the active principles responsible for the anticonvulsant activity of *Plumbago zeylanica*; flavonoidal compounds, present in this plant may likely be involved in this action. Hence this drug may be able to modulate the function of GABA or glutamate receptors

5. CONCLUSION

The anticonvulsant potential observed in the leaf galls and flower extract of *Plumbago zeylanica* against chemically induced convulsions in experimental animals underscores its significance in Ayurvedic and modern drug development. The presence of compounds like quercetin, kaempferol, flavonoidal compounds, and fixed oils is implicated in the plant's phyto medicinal uses.

REFERENCES

1. Ghani A, Medicinal plants of Bangladesh. Asiatic society of Bangladesh, Dhaka, Bangladesh, 1998, pp. 270.
2. Vijver LM et al. [1971] Antibacterial Activity in Roots of *Plumbago zeylanica* Planta Med. 20: 8-13.
3. Samineni R, Samathoti P, Gouru S.A, Khan A, Priyadharshini S.P, Manda K, Kishore M.V, Podila N. In-silico Investigation and Development of Cyclooxygenase-2 (1CX2) Selective Inhibition as a Possible Anti-inflammatory Activity. Biomed Pharmacol J 2024;17(3).
4. Aditi G et al. [1999]. Medicinal plants used in traditional medicine in Jimma zone, South West Ethiopia Pharm. Biol. 37: 321-323.
5. Vishnukanta et al. [2010] Evaluation of anti convulsant activity of *Plumbago zeylanica* Linn leaf extract Asian Journal of Pharmaceutical and Clinical Research 3(1): 76-78.
6. Lubaina A.S. et al. [2011] Shoot multiplication and direct organogenesis of an important medicinal plant *Plumbago zeylanica* L. (Plumbaginaceae). Journal of Research in Biology 6: 424-428.
7. Kirtikar KR and Basu BD, Indian medicinal plants. 2nd ed., Dehradun publisher Ltd, India, 1994, 1, pp. 830-832. 3. Hartwell JL, Plants used against cancer. A survey Lloydia, 34: (1971). 30-34.
8. Matangi suryaprabha, pharmacological evaluation of poly herbal extracts of *Amorphophallus paeoniifolius*, *citrus sinensis* and *Plumbago zeylanica* for antifungal activity. International journal of pharmacy and biological sciences 2018,8(4), 1261-1264.
9. Matangi suryaprabha, pharmacological evaluation of poly herbal extracts of *Amorphophallus paeoniifolius*, *citrus sinensis* and *Plumbago zeylanica* for antimicrobial and anthelmintic activity. activity. International journal of pharmacy and biological sciences 2019,9(1), 1421-1425.
10. Hang Chen 1,2, Juan Liu1, Kai Cui1, Qin Lu1, Chao Wang1,3, Haixia Wu1, Zixiang Yang1, Weifeng Ding 1, Shuxia Shao1, Haiying Wang1, Xiaofei Ling1, Kirst King- Jones4 & Xiaoming Chen1, Molecular mechanisms of tannin accumulation in *Rhus* galls and genes involved in plant-insect interactions, scientific reports, june 2018, 1-12.
11. Terrill T, Rowan AM, Douglas GB, Barry TN. Determination of extractable and bound condensed tannin concentrations in forage plants, protein concentrate meals and cereal grains. J Sci Food Agric. (2010) 58:321–329.
12. Thomas M.L.R.M.G., Filho J.M.B. Anti-inflammatory actions of tannins isolated from the bark of *Anacardium occidentale* L. J. Ethnopharmacol. 1985; 13:289–300. doi: 10.1016/0378-8741(85)90074-1.
13. Wafa N., Sofiane G., Mouhamed K. The antioxidant and antimicrobial activities of flavonoids and tannins extracted from *Phlomis bovei* De Noé Pelagia Res. Libr. Eur. J. Exp. Biol. 2016;6: 55–61.
14. Yazdi A., Sardari S., Sayyah M., Ezzati M.H. Evaluation of the anticonvulsant activity of the leaves of *glycyrrhiza glabra* var. *glandulifera* grown in Iran, as a possible renewable source for anticonvulsant compounds. Iran. J. Pharm. Res. 2011; 10:75–82.
15. Srinivasan K, Muruganandan S and Lal J, Evaluation of anti-inflammatory activity of *Pongamia pinnata* leaves in rats. J Ethnopharmacol, (2001). 78:151–157.
16. Mahli SS, Basu SP, Sinha KP and Banerjee NC, Pharmacological effects of karanjin and pongamol [from seed oil of *Pongamia pinnata*]. Ind J of Animal Science, 59: 657- 660.
17. Khosla P and Pandhi P, Anticonvulsant effect of nimodipin alone and in combination with diazepam on PTZ induced convulsion, Ind. J Pharmacol, (1989). 33: 208-211, (2001).
18. Shaik Khadar, Y., Kurni, L. D., BN, L. D., Sajida, A., Sibbala, S., Banothu, B., Gudipati, M., Kanakaraju, V. K., Podila, N. Design, Synthesis, Biological and Docking Studies of Novel 6-fluorobenzothiazole

- Substituted 1,2,4-Triazole Analogues as Prospective Anti-inflammatory Agents. *Egyptian Journal of Chemistry*, 2024; 67(12): 97-109.
19. Loscher W, Fassbender CP, and Noting B, The role of technical, biological and Pharmacological evaluation of anticonvulsant drugs in maximal electroshock seizure models, *Epilepsy Res*, (1991), 8:79-94.
 20. Kulkarni SK, Hand Book of Experimental Pharmacology, 2nd Edn. Vallab Prakashan, New Delhi 1993, pp. 78. 11. Ihan A, Iraz M, Kamisli S and Yigitoglu R, Pentylene tetrazole induced kindling seizure attenuated by Ginkgo biloba extract in mice, *Progress in Neuro-Psychopharmacology & biology Psychiatry*, 2006, pp. 78-84.
 21. Rajnarayan K, Sripal Reddy M, Chailuvadi MR and Krishna DR, Bioflavanoids classification, pharmacological, biochemical effect and therapeutic potential, *Ind J Pharmacol*, (2001), 32: 2-16.
 22. Kumar R, Kumar S, Patra A, Jayalakshmi S. Hepatoprotective activity of aerial parts of *Plumbago zeylanica* linn against carbon tetrachloride-induced hepatotoxicity in rats *Int J. Pharma Pharma Sci.*, 2009; 1:171-175.
 23. Hounnon A, Adomou AC, Gosling WD, Adeonipekun PA. 2021. A checklist of vascular plants of Ewe-Adakplame Relic Forest in Benin, West Africa. *PhytoKeys.*, 175: 151–174.
 24. Dubale S, Kebebe D, Zeynudin A, Abdissa N, Suleman S. Phytochemical Screening and Antimicrobial Activity Evaluation of Selected Medicinal Plants in Ethiopia. *J Exp Pharmacol*. 2023 Feb 8; 15:51-62.
 25. Samkeliso Takaidza, Fanyana Mtunzi, Michael Pillay, Analysis of the phytochemical contents and antioxidant activities of crude extracts from *Tulbaghia* species, *Journal of Traditional Chinese Medicine*. 38, 2018, 272-279.
 26. Babita Shukla, Poonam Kushwaha. Phytochemical screening and Antioxidant activity of roots of *Plumbago Zeylanica* L. *Current Indian Science*. 1,2023, 8.
 27. S. Brijesh, P.G. Daswani, P. Tetali, S.R. Rojatkari, N.H. Antia and T.J. Birdi, Studies on *Pongamia pinnata* (L.) Pierre leaves: understanding the mechanism(s) of action in infectious diarrhea, *J Zhejiang Univ Sci B*, (2006).7(8): 665–674.
 28. Prize Presentations: ACHARI PRIZE. *Indian J Pharmacol*. 2021 Dec;53(Suppl 1):S192–387.
 29. Al-Snafi AE. Medicinal plants with central nervous effects (part 2): plant based review. *IOSR Journal of Pharmacy* 2016; 6(8): 52-75.
 30. Naresh Podila, Penddinti NK, Rudrapal M, Rakshit G, Konidala SK, Pulusu VS, Bhandare RR, Shaik AB. Design, synthesis, biological and computational screening of novel pyridine-based thiadiazole derivatives as prospective anti-inflammatory agents. *Heliyon*. 2024, 30; 10(8): e29390.
 31. Podila Naresh, Akey KrishnaSwaroop, EsakkimuthukumarM, Shyam Sundar Pottabathula, Amarnath GS, Thirumoorthy A, Dharmaraja, S, Jubie S. Identification of plant secondary metabolites as dengue envelope protein entry inhibitors through an *in-silico* approach. *Journal of Medical Pharmaceutical and allied Sciences*. 2022, 11 (14), 5085 – 5095.
 32. P. Naresh, Pritosh Pattanaik, R.L. Priyadarshani, D. RaviSankar Reddy. Synthetic characterization and antimicrobial screening of some novel 6-fluoro benzothiazole substituted- 1, 2, 4 Triazoles analogues. *International Journal of Pharmacy & Technology*. 2013, 1 (1), 18-25.