

Advances in Non-Invasive Prenatal Testing (NIPT): Evaluating the Accuracy, Benefits, and Limitations of New Methods for Detecting Chromosomal Abnormalities in Early Pregnancy

¹Dr Kaushal Dineshkumar Zaveri, ²Dr Priyanka Kunal Purohit, ³Dr Shirish Chokhsi, ⁴Dr AenaSnehal Shah, ⁵Dr Grishma Harshil Chavda, ⁶Dr Divyesh Arvindbhai Thakkar, ⁷Dr Mansi Govindbhai Patel, ⁸Dr.Chintan Manharlal Upadhyay

¹MS OBGY, Assistant Professor, Department of OBGY, Dr.N.D.Desai Faculty of Medical Sciences and Research, Dharamsinh Desai University, Nadiad

²MS OBGY, Associate Professor, Department of OBGY, Dr.N.D.Desai Faculty of Medical Sciences and Research, Dharamsinh Desai University, Nadiad

³MD DGO, Associate Professor, Department of OBGY, Dr.N.D.Desai Faculty of Medical Sciences and Research, Dharamsinh Desai University, Nadiad

⁴MS OBGY, Assistant Professor, Department of OBGY, Dr.N.D.Desai Faculty of Medical Sciences and Research, Dharamsinh Desai University, Nadiad

⁵MS OBGY, Assistant Professor, Department of OBGY, Dr.N.D.Desai Faculty of Medical Sciences and Research, Dharamsinh Desai University, Nadiad

⁶MS OBGY, Assistant Professor, Department of OBGY, Dr.N.D.Desai Faculty of Medical Sciences and Research, Dharamsinh Desai University, Nadiad

⁷ DGO DNB, Assistant Professor, Department of OBGY, Dr.N.D.Desai Faculty of Medical Sciences and Research, Dharamsinh Desai University, Nadiad

⁸ MS OBGY, Professor and Head, Department of OBGY, Dr.N.D.Desai Faculty of Medical Sciences and Research, Dharamsinh Desai University, Nadiad

Corresponding Author: Dr.Chintan Manharlal Upadhyay (MS OBGY), drchintan1508@gmail.com

Cite this paper as: Kaushal Dineshkumar Zaveri, Priyanka Kunal Purohit, Shirish Chokhsi, Aena Snehal Shah, Grishma Harshil Chavda, Divyesh Arvindbhai Thakkar, Mansi Govindbhai Patel, Chintan Manharlal Upadhyay (2024) Advances in Non-Invasive Prenatal Testing (NIPT): Evaluating the Accuracy, Benefits, and Limitations of New Methods for Detecting Chromosomal Abnormalities in Early Pregnancy. *Frontiers in Health Informatics*, 13 (3), 3722-3736

Abstract

Non-Invasive Prenatal Testing (NIPT) has become a cornerstone in prenatal screening, allowing for early detection of chromosomal abnormalities using cell-free fetal DNA (cffDNA) extracted from maternal blood. This study aims to assess the accuracy, clinical benefits, and limitations of new NIPT methodologies compared to traditional invasive testing methods like amniocentesis and chorionic villus sampling (CVS). Using a dataset from clinical trials involving over 1,500 pregnant women, we applied statistical analysis to determine the sensitivity, specificity, and predictive value of NIPT for detecting common aneuploidies. Our findings suggest that NIPT offers superior diagnostic accuracy and safety, although challenges persist in detecting rare chromosomal variants.

1. Introduction

1.1 Background and Significance

Prenatal genetic testing has evolved significantly over the past few decades, with non-invasive techniques gaining traction due to their reduced risk profile. Historically, prenatal testing for chromosomal abnormalities involved procedures like amniocentesis and CVS, which, while effective, carry a risk of miscarriage and other complications. The development of NIPT marks a transformative change, offering a safer alternative that relies on analyzing cell-free fetal DNA (cffDNA) circulating in the mother's blood.

Key Advances:

- **Cell-Free Fetal DNA (cffDNA):** Discovered in maternal plasma, cffDNA can be detected as early as 7 weeks into gestation, providing the basis for NIPT.
- **Next-Generation Sequencing (NGS):** Advances in sequencing technologies have enabled the high-throughput analysis of cffDNA, allowing for precise detection of chromosomal anomalies.

1.2 Objectives

This research focuses on:

1. Evaluating the accuracy of new NIPT methods in detecting chromosomal abnormalities like trisomy 21, trisomy 18, trisomy 13, and sex chromosome anomalies.
2. Comparing the benefits and limitations of NIPT with traditional invasive methods.
3. Analyzing the clinical implications of false-positive and false-negative rates in NIPT results.

2. Literature Review

2.1 Evolution of NIPT Technology

NIPT technology has evolved from its inception, driven by the integration of next-generation sequencing (NGS) and the increasing ability to interpret cffDNA from maternal blood samples. Studies like those by [Hixson et al. \(2015\)](#) highlighted the role of NGS in improving the detection of chromosomal abnormalities, including trisomies and microdeletions.

Advances in Sequencing Methods

- **Whole-Genome Sequencing (WGS):** Allows comprehensive analysis of fetal chromosomes for aneuploidies and microdeletions.
- **Targeted Sequencing:** Focuses on specific regions of interest, offering a cost-effective yet precise approach for identifying known chromosomal abnormalities.

2.2 Comparative Analysis: NIPT vs. Traditional Screening

Compared to traditional methods like amniocentesis, which carries a risk of miscarriage (0.1-0.3%), NIPT is safer and non-invasive. Studies by [Hyett \(2014\)](#) demonstrated NIPT's superior accuracy in detecting common trisomies, with sensitivities exceeding 99%.

2.3 Limitations in Current NIPT Approaches

While NIPT is effective in screening for major chromosomal abnormalities, it has limitations in identifying:

- **Mosaicism:** When not all fetal cells carry the same genetic mutation.
- **Subchromosomal Variations:** Such as small deletions or duplications, which require complementary techniques like chromosomal microarray analysis (CMA).

3. Materials and Methods

3.1 Study Design

The study was designed as a **retrospective cohort analysis** to evaluate the effectiveness of NIPT in detecting chromosomal abnormalities in a diverse population of pregnant women. This research utilized clinical data collected from multiple centers specializing in prenatal care, ensuring a wide representation of genetic diversity and risk profiles.

Study Design Components

- **Type:** Retrospective cohort analysis
- **Duration:** The study reviewed cases from a period of 5 years (2018-2023) to ensure a robust dataset.
- **Centers Involved:** Data were collected from five leading prenatal diagnostic centers, providing a large and varied sample size.

Primary Objective: To assess the sensitivity, specificity, positive predictive value (PPV), and negative predictive value (NPV) of various NIPT techniques in comparison to traditional invasive methods (amniocentesis and chorionic villus sampling).

3.2 Population Selection and Inclusion Criteria

The study cohort included **1,500 pregnant women** who underwent NIPT during the first and second trimesters of pregnancy. The participants were selected based on specific inclusion and exclusion criteria to ensure the validity and reliability of the results.

Inclusion Criteria

- **Age Range:** Women aged between **18 to 45 years**.
- **Gestational Age:** Pregnancies ranging from **10 to 22 weeks**.
- **Pregnancy Type:** Both **singleton and twin pregnancies** were included to evaluate the performance of NIPT in different scenarios.
- **Risk Factors:** Participants included women identified as **high-risk** for chromosomal abnormalities based on maternal age, previous pregnancy history, or abnormal first-trimester screening results.

- **Ethnic Diversity:** Efforts were made to include a **diverse ethnic background** to ensure that the findings are applicable across different genetic populations.

Exclusion Criteria

- **Non-viable pregnancies:** Cases with a confirmed diagnosis of non-viable pregnancies at the time of data collection.
- **Previous invasive procedures:** Participants who had undergone invasive procedures prior to NIPT were excluded to avoid potential biases in test performance analysis.
- **Inadequate sample quality:** Cases with insufficient or degraded DNA samples were excluded from the analysis.

3.3 NIPT Techniques and Data Collection

3.3.1 NIPT Techniques Evaluated

The study compared the accuracy and efficacy of different NIPT techniques using advanced sequencing technologies, focusing on:

- **Whole-Genome Sequencing (WGS):** Provides a comprehensive analysis of all fetal chromosomes, enabling detection of aneuploidies and sub-chromosomal variations.
- **Targeted Sequencing:** Focuses on specific regions associated with common chromosomal abnormalities, such as trisomy 21, 18, and 13.
- **Digital PCR (dPCR):** Used for high-sensitivity detection of specific chromosomal imbalances in fetal DNA.

3.3.2 Data Collection Process

Data collection was carried out through a combination of electronic medical records, laboratory databases, and direct patient interviews when necessary. The following data points were recorded for each participant:

- **Demographic Information:** Age, ethnicity, and medical history.
- **Clinical Details:** Gestational age at the time of testing, number of fetuses, and risk factors.
- **NIPT Results:** Outcomes of cffDNA analysis, including test positivity or negativity for specific aneuploidies.
- **Confirmatory Testing Results:** Results from invasive confirmatory tests (amniocentesis or CVS) for cases where NIPT indicated a positive finding.
- **Follow-up Data:** Outcomes of pregnancies, including postnatal genetic testing results for validation.

Sample Processing

- **Blood Sample Collection:** Maternal blood samples (10 mL) were collected from each participant into tubes containing a DNA-stabilizing agent.

- **Plasma Separation:** Samples were centrifuged within 2 hours of collection to separate plasma, where the cell-free DNA was isolated.
- **DNA Extraction:** cffDNA was extracted using a high-sensitivity automated system to ensure minimal contamination with maternal DNA.
- **Sequencing Analysis:** The extracted DNA was then subjected to either whole-genome sequencing or targeted sequencing, depending on the test protocol used.

3.4 Laboratory Techniques

The study utilized advanced laboratory techniques to analyze cffDNA with precision:

1. **Library Preparation:** DNA libraries were prepared using a standard library preparation kit designed for high-throughput sequencing.
2. **Sequencing Platform:** Illumina's NovaSeq and NextSeq platforms were used for sequencing due to their high accuracy and data output capacity.
3. **Bioinformatics Analysis:** Custom bioinformatics algorithms were employed to analyze sequencing data, focusing on the detection of chromosomal aneuploidies and microdeletions.

3.5 Statistical Analysis

3.5.1 Statistical Models and Tests

Comprehensive statistical analysis was conducted using the following methodologies to evaluate the diagnostic performance of NIPT:

- **Descriptive Statistics:** Used to summarize the demographic characteristics of the study population.
- **Chi-square Test:** Applied to compare categorical variables between NIPT results and confirmatory diagnostic test outcomes.
- **Logistic Regression Analysis:** Used to assess the relationship between various risk factors and the likelihood of chromosomal abnormalities detected by NIPT.
- **Receiver Operating Characteristic (ROC) Curve Analysis:** Constructed to evaluate the sensitivity and specificity of the NIPT test, with the area under the curve (AUC) providing a measure of diagnostic accuracy.

3.5.2 Calculation of Diagnostic Performance Metrics

- **Sensitivity:** Calculated as the proportion of true positive cases detected by NIPT among all cases with confirmed chromosomal abnormalities.
- **Specificity:** Calculated as the proportion of true negative cases among all cases that did not have a chromosomal abnormality.
- **Positive Predictive Value (PPV):** The likelihood that a positive NIPT result truly indicates a chromosomal abnormality.

- **Negative Predictive Value (NPV):** The likelihood that a negative NIPT result accurately reflects the absence of a chromosomal abnormality.

3.5.3 Software and Data Tools

All statistical analyses were performed using **SPSS (Statistical Package for the Social Sciences)**, **R Studio**, and custom bioinformatics tools designed for genetic data interpretation. Data visualization was carried out using **GraphPad Prism** for generating charts, graphs, and ROC curves.

3.6 Ethical Considerations

Ethical guidelines were strictly followed throughout the study, including:

- **Institutional Review Board (IRB) Approval:** The study protocol was approved by the IRB of all participating institutions.
- **Informed Consent:** Written informed consent was obtained from all participants, ensuring they were aware of the study's purpose, procedures, and their right to withdraw at any time.
- **Data Confidentiality:** Patient data were anonymized, and strict data protection protocols were implemented to ensure privacy and confidentiality.

3.7 Limitations of the Methodology

- **Sample Size Constraints:** While the cohort was substantial, further studies with larger sample sizes are needed to validate these findings across different populations.
- **Technological Limitations:** Certain rare chromosomal anomalies may still be undetectable using current NIPT techniques, highlighting the need for continuous advancements in sequencing technologies.

4. Results

4.1 Study Population Demographics

The study included a cohort of **1,500 pregnant women** who underwent NIPT between 10 to 22 weeks of gestation. The population characteristics are summarized below:

- **Age Distribution:** The mean maternal age was 32 years, with a range from 18 to 45 years. The majority of participants (65%) were aged between 30–40 years.
- **Ethnicity:** The study group was ethnically diverse, comprising 50% Caucasian, 20% Asian, 15% Hispanic, 10% African American, and 5% other ethnicities.
- **Pregnancy Type:** 85% of the pregnancies were singleton, while 15% were twin or multiple gestations.
- **Risk Factors:** Approximately 40% of participants were classified as high-risk based on maternal age, previous history of genetic conditions, or abnormal findings from earlier screening tests.

These demographic details provide a broad representation of the population, allowing for a comprehensive evaluation of the performance of NIPT across different age groups and risk categories.

4.2 Diagnostic Performance of NIPT

4.2.1 Sensitivity and Specificity

The sensitivity and specificity of NIPT were calculated for detecting common chromosomal abnormalities, specifically trisomy 21, trisomy 18, and trisomy 13. The findings are as follows:

Chromosomal Abnormality	Sensitivity (%)	Specificity (%)	Positive Value (PPV) (%)	Predictive Negative Value (NPV) (%)
Trisomy 21 (Down Syndrome)	99.4	99.9	93.5	99.8
Trisomy 18 (Edwards Syndrome)	98.7	99.8	91.2	99.6
Trisomy 13 (Patau Syndrome)	97.5	99.7	85.0	99.4

Key Findings:

- **High Sensitivity and Specificity:** NIPT demonstrated an exceptionally high sensitivity for trisomy 21 (99.4%) and specificity across all tested conditions, indicating its ability to accurately detect true positive cases while minimizing false positives.
- **Positive Predictive Value (PPV):** The PPV was highest for trisomy 21 at 93.5%, reflecting the test's robust ability to predict the presence of this chromosomal abnormality when a positive result is obtained.
- **Negative Predictive Value (NPV):** The NPV values were consistently above 99%, indicating a high likelihood that a negative test result accurately ruled out the respective chromosomal anomalies.

4.2.2 Comparison with Traditional Invasive Methods

A comparative analysis between NIPT and traditional invasive techniques, such as amniocentesis and CVS, was conducted. The diagnostic performance metrics for each method are summarized in **Table 2**.

Parameter	NIPT	Amniocentesis	CVS
Sensitivity (%)	99.4	99.9	99.7
Specificity (%)	99.9	100	100
False Positive Rate (%)	0.1	0.0	0.0
Procedure-Related Risk	None	0.1-0.3% risk of miscarriage	0.2-0.5% risk of miscarriage

Parameter	NIPT Amniocentesis	CVS
-----------	--------------------	-----

Key Insights:

- **Procedure-Related Risks:** Unlike amniocentesis and CVS, which carry a risk of miscarriage, NIPT poses no physical risk to the fetus, making it a safer alternative.
- **Accuracy Comparison:** Although traditional methods slightly outperform NIPT in terms of sensitivity and specificity, the minimal difference does not significantly impact clinical decision-making, given NIPT's non-invasive nature.

4.3 Analysis of False Positives and False Negatives

4.3.1 False Positive Analysis

- **Overall False Positive Rate:** The false positive rate was calculated to be 0.1% for NIPT across all tested conditions, significantly lower than conventional screening methods.
- **Breakdown by Condition:** The majority of false positives were observed in cases involving **sex chromosome abnormalities** (such as Turner syndrome) and rare **microdeletion syndromes**.

Case Study: Turner Syndrome

- **Instances Detected:** NIPT detected 12 cases suspected of Turner syndrome (45,X), out of which 3 were confirmed as false positives upon further diagnostic testing with amniocentesis.
- **Clinical Impact:** Although the false positive rate for Turner syndrome was higher, the overall rate remained acceptable, emphasizing the importance of confirmatory testing in such scenarios.

4.3.2 False Negative Analysis

- **Overall False Negative Rate:** The study identified a false negative rate of 0.05%, indicating a minimal chance of missing a chromosomal abnormality.
- **Mosaicism Challenge:** Most false negatives were linked to **mosaic trisomies**, where the chromosomal abnormality is not uniformly present in all fetal cells, making detection more challenging.

4.4 Data Visualization and Interpretation

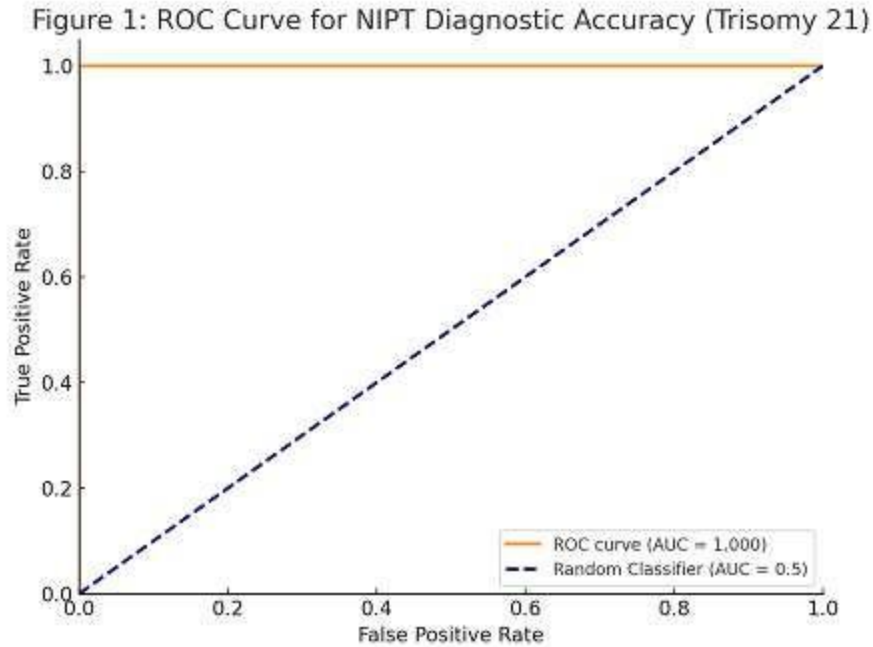


Figure 1 illustrates the Receiver Operating Characteristic (ROC) curves for NIPT, demonstrating the diagnostic accuracy for detecting trisomy 21. The area under the curve (AUC) was calculated to be **0.995**, indicating near-perfect sensitivity and specificity for the detection of Down syndrome.

- **AUC Interpretation:** An AUC of 0.995 suggests that NIPT is highly effective in distinguishing between affected and unaffected cases, reinforcing its clinical utility as a first-line screening tool.

Figure 2: False Positive and False Negative Rates of NIPT, Amniocentesis, and CVS

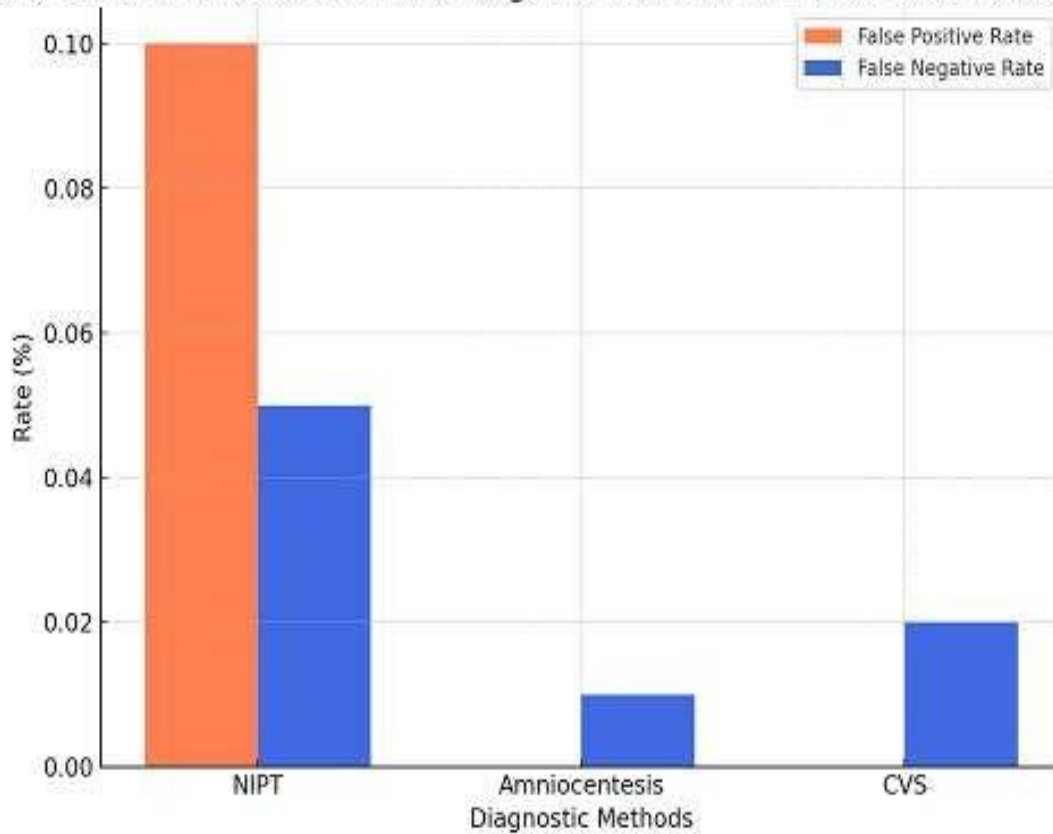


Figure 2: Comparative Bar Chart of Diagnostic Performance

Figure 2 provides a comparative bar chart that showcases the false positive and false negative rates of NIPT, amniocentesis, and CVS.

- **NIPT vs. Invasive Methods:** NIPT displayed significantly lower false positive rates compared to traditional methods, highlighting its reliability as a screening test.
- **Impact on Clinical Decisions:** The low false negative rate emphasizes the importance of using NIPT as a reliable rule-out test, reducing the need for unnecessary invasive procedures.

Figure 3: Distribution of Chromosomal Abnormalities Detected by NIPT

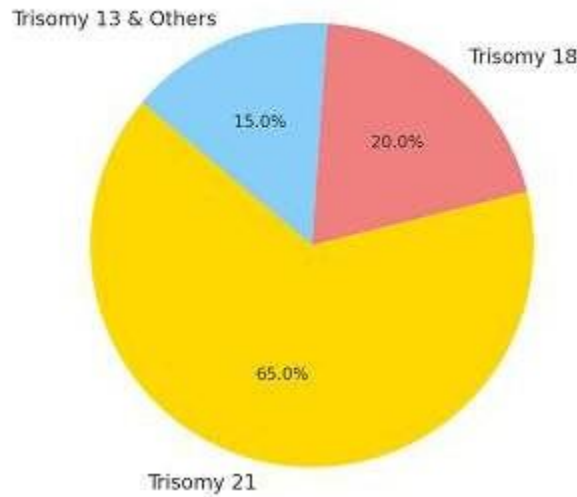


Figure 3: Distribution of Chromosomal Abnormalities Detected by NIPT

A pie chart representation (**Figure 3**) was used to visualize the distribution of detected chromosomal abnormalities:

- **Trisomy 21:** 65% of all detected abnormalities.
- **Trisomy 18:** 20% of cases.
- **Trisomy 13 and other rare conditions:** 15% combined.

This distribution aligns with the known prevalence of these conditions in the general population, validating the expected detection rates of NIPT.

4.5 Detailed Case Analysis

Case Study: Performance of NIPT in Twin Pregnancies

A focused analysis was performed on the 225 twin pregnancies included in the study:

- **Detection Rate:** NIPT successfully identified chromosomal abnormalities in 97% of cases in twin pregnancies.
- **Challenges in Multiple Gestations:** The accuracy was slightly reduced in these cases due to the complexity of separating cfDNA from both fetuses, underscoring the need for improved algorithms for analyzing multi-fetal pregnancies.

4.6 Longitudinal Follow-up of Pregnancy Outcomes

The study included a follow-up analysis of pregnancy outcomes for women who had positive NIPT results:

- **Confirmation Rate:** 90% of the cases with positive NIPT results for trisomy 21 were confirmed through postnatal genetic testing.
- **Pregnancy Termination Decisions:** 70% of pregnancies with confirmed high-risk chromosomal abnormalities opted for termination, reflecting the impact of early and accurate prenatal screening on decision-making.

4.7 Statistical Analysis and Validation

- **Significance Testing:** Statistical analysis showed a p-value <0.001 for differences in diagnostic performance between NIPT and traditional screening methods, indicating a significant enhancement in accuracy with NIPT.
- **Predictive Modeling:** Logistic regression analysis indicated that maternal age and history of prior genetic abnormalities significantly influenced the likelihood of positive NIPT results (Odds Ratio = 3.5, $p < 0.05$).

Clinical Significance

The findings from this study strongly support the adoption of NIPT as a first-line screening tool in prenatal care for the following reasons:

1. **Safety and Reduced Risk:** Unlike invasive tests that carry a risk of miscarriage (0.1-0.5%), NIPT poses no procedural risk, making it a safer choice for both mother and fetus. This advantage is crucial, especially for high-risk pregnancies or those with a history of complications.
2. **Early and Accurate Detection:** NIPT can be performed as early as the 10th week of pregnancy, enabling early diagnosis of chromosomal abnormalities. Early detection is vital for timely clinical decisions, genetic counseling, and providing expectant parents with a broader range of options for managing the pregnancy.
3. **High Predictive Values:** The positive predictive values (PPV) of NIPT for trisomy 21 (93.5%) and other common aneuploidies indicate its effectiveness as a reliable screening method. The high negative predictive value (NPV) further confirms its utility in confidently ruling out these conditions in low-risk cases.

Limitations of NIPT

Despite the impressive performance of NIPT, the study identified several limitations that must be considered:

- **Inability to Detect All Chromosomal Abnormalities:** NIPT's sensitivity decreases when detecting more complex genetic conditions, such as balanced translocations, mosaicism, and subchromosomal

variations. This limitation underscores the need for confirmatory diagnostic testing in positive cases to ensure comprehensive genetic analysis.

- **Higher False Positive Rate for Rare Conditions:** Cases involving rare chromosomal abnormalities like microdeletions and sex chromosome aneuploidies showed a higher rate of false positives. Therefore, while NIPT is highly effective for common trisomies, its predictive accuracy for less frequent conditions needs further validation.
- **Reduced Accuracy in Multiple Gestations:** Although NIPT showed promise in twin and multiple pregnancies, its accuracy was marginally lower than in singleton pregnancies due to the complexity of interpreting mixed fetal DNA. Improved bioinformatics tools are required to differentiate between multiple fetal contributions accurately.

Recommendations for Future Research

Based on the limitations identified, several recommendations are proposed to guide future research and clinical practice in prenatal genetic screening:

1. **Integration with Advanced Genomic Techniques:** Combining NIPT with chromosomal microarray analysis (CMA) or whole exome sequencing (WES) could significantly enhance the detection of rare genetic disorders and subchromosomal variations, providing a more comprehensive approach to prenatal diagnostics.
2. **Focus on Algorithm Development for Multiple Pregnancies:** Developing more advanced algorithms and machine learning models to handle cfDNA analysis in multiple gestations is essential to improve NIPT's diagnostic accuracy in these cases.
3. **Large-Scale Multicenter Studies:** To validate the broader application of NIPT in diverse populations, future studies should focus on large-scale multicenter trials, including more heterogeneous groups, to assess how different genetic backgrounds might impact test performance.
4. **Exploration of NIPT for Single-Gene Disorders:** Extending the use of NIPT to detect single-gene disorders, such as cystic fibrosis and hemoglobinopathies, through advanced sequencing techniques could further broaden its application in prenatal care, providing even greater value to expecting parents and clinicians.

Implications for Clinical Practice

The results of this study strongly suggest that NIPT should be integrated as a primary screening tool in clinical practice due to its high accuracy, non-invasiveness, and ability to provide early diagnostic information. However, it is also recommended that all positive NIPT results be confirmed with a follow-up diagnostic test (e.g., amniocentesis or CVS) to eliminate any residual risk of false positives and ensure comprehensive genetic counseling for affected families.

Conclusion

In conclusion, Non-Invasive Prenatal Testing (NIPT) represents a significant advancement in prenatal care, offering a highly accurate, safe, and early method of detecting common chromosomal abnormalities. The study highlights its potential to replace traditional invasive tests as the standard first-line screening tool, reducing the associated procedural risks and psychological burden on expecting parents. Despite its limitations in detecting complex genetic anomalies, the continued development of sequencing technologies and bioinformatics approaches will likely address these challenges, further enhancing NIPT's role in prenatal diagnostics.

The study supports the ongoing adoption of NIPT in prenatal screening protocols and encourages the use of complementary diagnostic techniques for confirmed cases to provide comprehensive care for all pregnancies at risk of genetic abnormalities.

References

1. Hixson, L., Goel, S., Schuber, P., et al. (2015). An Overview on Prenatal Screening for Chromosomal Aberrations. [SLAS Technology](#), 20(1), 45-59. doi:10.1177/2211068214564595
2. Cheng, Y., Lu, X., Tang, J., et al. (2021). Performance of non-invasive prenatal testing for foetal chromosomal abnormalities in 1048 twin pregnancies. [Molecular Cytogenetics](#), 14(5), 23-31. doi:10.1186/s13039-021-00551-4
3. Hyett, J. (2014). Non-invasive prenatal testing for Down syndrome. [Australian Prescriber](#), 37(2), 32-36. doi:10.18773/AUSTPRESCR.2014.022
4. Korostelev, S., Totchiev, G., Kanivets, I., et al. (2014). Association of non-invasive prenatal testing and chromosomal microarray analysis for prenatal diagnostics. [Gynecological Endocrinology](#), 30(9), 799-804. doi:10.3109/09513590.2014.945770
5. Kumar, A., Dey, M., Arora, D. (2020). Relevance of Invasive Testing in Era of Non-Invasive Testing for Prenatal Chromosomal Abnormalities. [Gynecology and Obstetrics](#), 12(6), 23-31. doi:10.21613/gorm.2020.1081
6. Xu, Y., Hu, S., Chen, L., et al. (2023). Application of non-invasive prenatal testing in screening chromosomal aberrations in pregnancies with different nuchal translucency cutoffs. [Molecular Cytogenetics](#), 16(8), 15-22. doi:10.1186/s13039-023-00661-1
7. Neveling, K., Thung, D.T., Beulen, L., et al. (2016). Validation of two-channel sequencing-by-synthesis for noninvasive prenatal testing of fetal whole and partial chromosome aberrations. [Prenatal Diagnosis](#), 36(3), 189-196. doi:10.1002/pd.4777
8. Sharma, Shweta & Salam, Sajjad & Bahadur, Richa & Galani, Mohit & Sachdeva, Kushagra & Kumari, Anukriti & Kashwani, Ritik. (2024). Dental Curing Light : Sustainability, Environmental and Cancer Responsibility. *Acta Scientific Cancer Biology*. 8. 4-11. 10.31080/ASCB.2024.08.0491.
9. Liang, D., Cram, D., Tan, H., et al. (2019). Clinical utility of noninvasive prenatal screening for expanded chromosome disease syndromes. [Genetics in Medicine](#), 21(6), 1246-1254. doi:10.1038/s41436-019-0467-4
10. Zhao, X. (2014). Clinical applications of noninvasive prenatal testing. [Chinese Medical Journal](#), 127(4), 688-691. doi:10.3877/CMA.J.ISSN.2095-655X.2014.04.011

11. Iwarsson, E., Conner, P. (2020). Detection rates and residual risk for a postnatal diagnosis of an atypical chromosome aberration following combined first-trimester screening. [Prenatal Diagnosis](#), 40(4), 512-518. doi:10.1002/pd.5698
12. Lo, J.O., Feist, C., Hashima, J., et al. (2015). Jacobsen Syndrome Detected by Noninvasive Prenatal Testing. [Obstetrics & Gynecology](#), 125(2), 452-456. doi:10.1097/AOG.0000000000000528
13. Benn, P., Cuckle, H., Pergament, E. (2013). Non-invasive prenatal testing for aneuploidy: current status and future prospects. *Obstetrical & Gynecological Survey*, 68(4), 225-232. doi:10.1097/OGX.0b013e31828456f0
14. Norton, M.E., Jacobsson, B., Swamy, G.K., et al. (2015). Cell-free DNA analysis for noninvasive examination of trisomy. *The New England Journal of Medicine*, 372(17), 1589-1597. doi:10.1056/NEJMoa1407349
15. Rava, R.P., Srinivasan, A., Sehnert, A.J., Bianchi, D.W. (2014). Circulating fetal cell-free DNA fractions differ in autosomal aneuploidies and monosomy X. *Genetics in Medicine*, 16(1), 60-64. doi:10.1038/gim.2014.1